Antifungal Activity of Nettle (Urtica dioica L.) and European Pennyroyal (Mentha pulegium L.) Extracts on Alternaria alternata

Mitra Sayidi 1 and Sevil Nematollahi* 2

1. Former MSc student, Department of Plant Protection, Malekan branch, Islamic Azad University, Malekan, Iran
2. Assistant Prof. Department of Plant Protection, Tabriz branch, Islamic Azad University, Tabriz, Iran

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ABSTRACT
Increasing public concerns about fungicide residues have prompted interests in biological control especially use of herb extracts derived from several medicinal plants as an alternative strategy for disease management. Recently tomato leaf spot disease caused by Alternaria alternata has become epidemic in almost areas of Iran. This study was conducted for the identification of the effect of antifungal activity of two medicinal plants extracts; Mentha pulegium L. and Urtica dioica L. against A. alternata. Both plant extracts exhibited antifungal potential against the pathogenic fungus. The Nettle extract showed a more degree of inhibition with 59.78% and 77.81% inhibition rate at concentration of 1000 and 1500 ppm, respectively. The inhibition of the mycelial growth was strongly reduced in the concentration of 1500 ppm of both plant extracts. The results of this study confirm that extracts of the selected medicinal plants can be used as an alternative fungicide in control of this pathogenic fungus.

1. Introduction

Various plant pathogens as fungi, viruses and bacteria reduce plant productivity and lead to huge losses (Tapwal et al., 2011). The Alternaria associated with a wide variety of substrates including seeds, plants, agricultural products, animals, soil and the atmosphere. Alternaria species are known as important plant pathogens which mostly cause diseases on aerial parts of many plants worldwide and causing major losses on a variety range of crops (Woudenberg, 2013). A. alternata usually reported as a opportunistic pathogen which attack vigorous plants and cause economic losses (Ellis, 1985). The A. alternata is a saprophytic pathogen of tomato causing postharvest losses (Akhtar, 1994).

One of the most efficient method for protecting the crops against fungal infection is the use of saprophytic pathogen of tomato causing postharvest losses fungicides (Akhtar, 1994).

However, these chemical compounds are toxic and easily transmit to non-target species even polluted the environment through air, water, soil etc. Its residual effects also persist in food chain for longer time (Tapwal, 2011). In recent years the increasing of resistance of pathogenic fungi towards the synthetic fungicides is one of great concern. Therefore, it has made necessary to use some biological approaches which have been advanced as potential alternatives to synthetic fungicides for the management of diseases (Wang, 2008).
Traditionally, many plants oils and extracts have been reported with antimicrobial properties. It is important to investigation for new plant-derived natural substances that can obstruct the fungal pathogenicity and is considered as a safe alternate to synthetic fungicides in plant disease managements, due to their less negative impact on environment (Gujar, 2012; Hadizadeh, 2009). Nowadays, there has been a growing interest in the determination of the biological properties of herb extracts derived from several medicinal plants such as Mentha and Urtica species. Mentha pulegium L. and Urtica dioica L. are considered as medicinal plants because of its pharmacological and biological properties (Motamedi, 2014; Joshi, 2014).

The objective of this study was the exploring the in vitro antifungal potential of plant extracts for inhibition activity against Alternaria alternata.

2. Materials and Methods

2.1. Collection of plant materials

Mentha pulegium L. and Urtica dioica L. plants used in this study were collected from the surrounding area of Harbil, Tabriz, Iran during 2014, and brought to the laboratory. Tomato plants were collected from agricultural lands in East Azarbaijan province.

2.2. Isolation of the pathogen

Fungal pathogen was isolated from infected tomato that collected from Tabriz province of Iran, during 2014. Samples transferred to mycology lab of Azad University of Malekan and stored at 4°C. Plant materials were washed in running tap water for 10 min. A small piece of leaf tissue (5×5 mm) was taken from the margin of a lesion, surface disinfected with 1% sodium hypochlorite solution for 1 min and then rinsed with sterile water (Frisvad 1983). Sterile small pieces were placed in Petri dishes containing potato dextrose agar (PDA), then incubated at 24±1°C until the pure fungal colonies were appeared. Pure culture was established via single spore and maintained on PCA (20 g white potato, 20 g carrot, and 20 g agar per 1 L of distilled water). The cultured fungus identified as A. alternata using morphological characteristics, according to the description provided by Simmons, 2007.

2.3. Preparation of plant extracts

Leaves of selected plants were thoroughly washed in running water and kept in shade to dry. Dry materials then grinded using a cutting mill. Pure methanol (96%) was used as solvent. Four hundred milliliter of solvent were added to 40 g of dry powder materials of each plant and homogenized for 1 h with a magnet. After two days’ mixtures were passed through 0.2 μ size Whatmann filter paper then shaken at 160 rpm and 40 C to obtain clear extracts. The methanol was completely removed from clear solutions using a rotary evaporator. Exact extract concentrations (500, 1000 and 1500 ppm) were prepared in methanol solvent (Zaker, 2010).

2.4. Antifungal evaluation of plant extracts on mycelial growth of A. alternata in vitro

In this experiment, the extracts were mixed with sterile molten PDA to obtain final concentrations 500, 1000 and 1500 ppm, and in the case of control extract free PDA is used. The PDA was poured into 80 mm Petri dishes and then inoculated with 4 mm plugs from 7-day-old culture of A. alternata. The fungal pathogen was placed in the center. Petri dishes were incubated at 25°C. The measurements of the mycelial growth dynamic of the fungus were recorded on a daily basis, beginning with 24 hours after inoculation. Four replicates were used per treatment and the screening procedure repeated twice. Percentage inhabitation of mycelia growth was calculated according to the formula (Mohana, 2007):

\[
\text{Percentage inhibition} = \frac{(C-T)}{C} \times 100
\]

Where C, average diameter of the fungal colony of the control plate and T, average diameter of the fungal colony treated with the treatment plate.

2.5. Statistical analysis

Statistical analysis of the data obtained in the present study was carried out in a completely randomized design layout using SPSS ver. 19, where the comparison of means of different treatment was performed using factorial design.
3. Results

In the present investigation, various concentrations of plant extracts significantly showed antifungal potential against *A. alternaria*, which is demonstrated by inhibition of fungal mycelia growth (Table 1). Table 2 shows means comparison of the effect of different concentrations the plant extracts on mycelial growth. The results revealed that the antifungal activity of the extracts being positively influenced by increasing the concentration of the extracts in the growth media. The plant extracts at 1500 ppm showed potent and inhibitory effect on the radial growth of *A. alternata*. The final measurements revealed that the extracts obtained from *Mentha pulegium* had highest inhibitory activity, while *Urtica dioica* presented lower inhibition activity against tested pathogen mycelium growth.

Based on the results, all extracts concentrations had significant inhibition on the growth of *A. alternata* (Fig1). ranging from 67.87 and 77.81% at 1500 ppm of *Urtica dioica* and *Mentha pulegium* extracts, respectively. The lowest antifungal activity of plant extracts results was related to the concentration of 500ppm which had 40% inhibition of growth approximately in both extracts.

### Table 1. Variance analysis of effects of plant extract for control of *A. alternata*.

<table>
<thead>
<tr>
<th>Source of variation</th>
<th>Degree of freedom</th>
<th>Means square</th>
<th>F</th>
</tr>
</thead>
<tbody>
<tr>
<td>Plant extracts</td>
<td>1</td>
<td>2.195</td>
<td>92.727**</td>
</tr>
<tr>
<td>Concentration</td>
<td>3</td>
<td>86.055</td>
<td>36.353**</td>
</tr>
<tr>
<td>Daily records</td>
<td>5</td>
<td>50.613</td>
<td>21.383**</td>
</tr>
<tr>
<td>Plant extracts × concentration</td>
<td>3</td>
<td>0.459</td>
<td>19.375**</td>
</tr>
<tr>
<td>Plant extracts × daily records</td>
<td>5</td>
<td>0.051</td>
<td>2.172 ns</td>
</tr>
<tr>
<td>Concentration × daily records</td>
<td>15</td>
<td>4.056</td>
<td>171.329**</td>
</tr>
<tr>
<td>Plants extracts × concentration × daily records</td>
<td>15</td>
<td>0.037</td>
<td>1.557 ns</td>
</tr>
<tr>
<td>Error</td>
<td>143</td>
<td>0.024</td>
<td></td>
</tr>
</tbody>
</table>

**Significant at 1% probability level.

### Table 2. Mean comparison of effects of plant extracts concentration on mycelial growth of *A. alternata*.

<table>
<thead>
<tr>
<th>Plants name</th>
<th>Mean mycelial growth</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0 ppm</td>
</tr>
<tr>
<td><em>Mentha pulegium</em></td>
<td>4.45 a</td>
</tr>
<tr>
<td><em>Urtica dioica</em></td>
<td>4.45 a</td>
</tr>
</tbody>
</table>

Means within each column followed by same letter are not significantly different at 1% probability level.
4. Discussion

Plant extract include ethanol extracts and its fractions, resin and essential oils have been reported to have antifungal activity (Gahukar, 2012). Most investigation of plant extracts have been shown effective inhibitory against a wide range of pathogens in vitro but these researches almost failed to control diseases under field conditions (Benner, 1993; Chang, 2008). However, in vitro screening of plant extracts is an important first step in identifying plants with potential application (Tegegne, 2007).

Previously, Tegegne (2008) showed that different natural substances, such as essential oils and extracts of different species have varying degree of growth inhibitory effects against some Alternaria, Fusarium, Botrytis and Rhizoctonia. Methanolic extracts of six selected plant leaves screened for their antifungal activity against A. alternata at 5, 10 and 20% concentration in vitro. Phytochemical analysis of leaf extracts determined the presence of some chemical compounds including Alkaloids, Terpenoids, Phenols, Saponins and Tannins at various concentrations (Singh, 2014).

Rodino (2014), showed in vitro effects of antifungal properties of ethanolic and aqueous extracts of traditional medicinal plants including; Xanthium strumarium, Artemisia absinthium, Rosmarinus officinalis and Datura stramonium, against plant pathogenic fungus Alternaria alternata. Also recently Buch and Arya (2017) showed that three main pathogenic fungal diseases in Cotton (Gossypium herbaceum) can be successfully controlled using selected plant extract and have demonstrated that plant extracts have potential capabilities for use as chemical fungicides replacement. Similar to this research, it has been reported that three plant extracts have significant inhibition effects on Alternaria solani (Tomazoni et al., 2017).

Although mechanism of action of plant extracts and essential oils is unclear, several studies have been conducted to understand their function. Several researches attributed this function to the phenolic compounds and some investigations have been proposed to possible action mechanisms by which mycelial growth may be reduced or totally inhibited. Researches showed that it is the toxic effects of essential oils components and extracts on the functionality and structure of the cell membrane (Sikkema, 1995; Veldhuizen, 2006). Essential oils components and extracts might act on the hyphae of the mycelium and resulting in collapse and death of the mycelium, the loss of rigidity and integrity of the hypha cell wall, by providing exit of components from the cytoplasm (Sharma, 2006).

Over the past two decades’ extensive investigations in an attempt to reduce the use of synthetic fungicides have been made into the possible exploitation of plant compounds as
natural commercial products that are safe for humans and the environment, and also they are being easily accessible and relatively cost effective (Daayf, 1995; Rodino, 2014).

In this study, the ability of ethanolic extracts derived from selected plants to inhibit the mycelial growth of A. alternata was evaluated in vitro contact assays. The active extract was Mentha pulegium followed by Urtica dioica. Antifungal screening indicated that Mentha pulegium and Urtica dioica extracts at a concentration of 1500 ppm markedly inhibited the mycelial growth of test pathogen.

In conclusion, examination of various concentrations of Mentha pulegium and Urtica dioica on A. alternata in this study showed promising prospects for the utilization of natural plants extracts in postharvest disease control. So organic extract of selected medicinal plants could be applied as alternative industrial products to synthetic fungicides for using in agro-industries and also to screen and develop such novel types of selective and natural fungicides in the biocontrol of many agricultural plant pathogens causing drastic losses to crop.

References


